



SKM VAIDHYA AMIRTHAM

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“भोजनान्ते पिबेत्तक्रं वासरान्ते पिबेत्पयः ।
निशान्ते च पिबेद्द्वारि त्रिभिर्रोगो न जायते ॥”



Drinking buttermilk at the end of a meal, milk at the end of the day,
and water in the morning upon waking helps maintain good health
and prevents diseases.

Articles are invited in Siddha, Ayurveda and Unani fields about clinical experience, rare medicinal preparations, successful treatments, Herbal informations and AYUSH Foods for our "SKM Vaidhya Amirtham" News letter which has around 10000 copies of circulation.

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Evaluation of Chronic toxicity of Urai mathirai - Siddha Herbal Formulation

INTRODUCTION:

Traditional Systems of medicines are playing a key role in meeting the global health care needs. India has different recognized systems of medicine. They are Ayurveda, Siddha, Unani, Yoga, Naturopathy, Homeopathy and Sowa Rigpa. Among them Siddha is the unique system of medicine which is originated from Tamil Nadu and has its origins in Tamil language. Literally the word “Siddha” means “established truth”. Siddha system of medicine was claimed to alleviate the root cause of the diseases by maintaining the ratio of Vatham, Pitham and Kapham. The origin of the Siddha system of medicine is attributed from ancient saint called Siddhars⁽¹⁾.



Urai Mathirai is a drug used for the past three decades in the form of long finger size bullets which are rubbed and administered through breast milk with children's to improve immunity to get free from health hazards such as frequent respiratory infections /gastrointestinal infections and anorexia. For global acceptance, this system of medicine should undergo scientific validation, i.e., upgrading the one of the levels is safety of the dose⁽²⁾.

The acute and sub acute toxicity of the Siddha herbal formulation urai mathirai was administered orally at a dose of 10, 50 and 100 mg/kg.b.wt for 28 days does not showed any toxicity effect in wistar rats.

Hence the present study was performed to evaluate the chronic toxicity of the Siddha herbal formulation Urai Mathirai in experimental wistar rats. Through this study the safety of this herbal drug can be established for the clinical use of this traditional formulation among the children's to improve immunity.



MATERIAL AND METHODS:

Test animal

Wistar albino rats of both sex (Male and female) weighing about 130–250 g were used in the chronic toxicology studies. The rats were housed in the Department of Pharmacology, Siddha central research institute, Chennai. The rats were kept in sanitized polypropylene cages housed with sterile corn cob as bedding materials at animal house, in an air conditioned environment with four rats in each cage and maintained at room temperature of $(23 \pm 2)^{\circ}\text{C}$ with relative humidity $(60\% \pm 10\%)$ under 12 hour dark and light cycle⁽³⁾. Rats were given free access to standard pellet diet and water ad libitum⁽⁴⁾. All experimental procedures were in compliance with the Committee for the Purpose of Control and Supervision of Experiments on animals (CPCSEA) and were approved by Institutional Animal Ethical Committee, with an approval number **162/Pharma/SCRI/2017**.

Dose calculation

The clinical dose of Urai mathirai in children is 50 mg (HED = 2.5mg/kg body weight). The animal (rat) doses are calculated as per the FDA guidelines and the calculated therapeutic dose (TD) was found to be 10mg/kg of body weight. In this study to determine the dose correlated effects, 5 times TD and 10 times TD i.e. 50 mg/kg & 100 mg/kg of bodyweight were chosen correspondingly.

Experimental design

The oral chronic toxicity study was evaluated accordance with to Organization for Economic Cooperation and Development (OECD) guideline 408 and Central Council for Research in Ayurvedic Sciences (CCRAS) guideline (6, 7). Wistar albino Rats of both sexes (over night fasting free access to water), aged 8–12 weeks old were used. In 80 rats (40 Male and 40 females) were randomized into groups separately based on bodyweight. After the randomization process, each study animal was assigned a unique number and identified by a picric acid mark. Group-I served with vehicle as normal control, Group-II, Group-III and Group-IV administered with test drug orally at a dose of 10 mg/kg, 50 mg/kg and 100 mg/kg respectively for 90 days. Before drug administration, the body weight of each animal was determined and the dose was calculated according to the body weight for every week consecutively for 90 days (8). Rat general health, and signs of toxicity, body weight, mortality, food and water intake was monitored. At 90th day 50% of the experimental animals (40 animals from both sex) in each group were subjected to euthanasia.



The necropsy was carried out on all euthanized animals and the organs were isolated and observed macroscopically for abnormalities. After 30 days of post treatment the remaining half animals (40 animals) were euthanized and organs were collected and observed macroscopically for abnormalities. The collected organs during the treatment and post treatment were preserved in 10% formaldehyde solution for histopathological examination.

Relative organ weight

The internal organs (brain, heart, thymus, lungs, liver, stomach, spleen, kidney, adrenal gland, pancreas and sex organs) excised from all the experimental rats after 90th day and 120th day. Organ-to-body weight ratio Posted on Authorea 23 Apr 2020 — CC BY 4.0 — <https://doi.org/10.22541/au.158765852.23761913> — This a preprint and has not been peer reviewed. Data may be preliminary. was calculated by dividing the weight (g) of each organ by the weight (g) of rats before sacrifice.

Biochemical parameters

The biochemical analysis were done on serum after centrifugation of collected blood and the following parameters like Blood glucose, Total Cholesterol, Triglyceride, HDL, LDL, SGOT, SGPT, ALP, GGT, Total Protein, Albumin, LDH, CRP, Creatinine Kinase levels, Creatinine Kinase – MB, Urea, Serum creatinine, Total Bilirubin, Uric acid, Serum Calcium were determined for both control and Urai mathirai treated groups by using standard biochemical method.

Haematological parameters

The haematological parameters such as white blood cell (WBC), red blood cell (RBC), lymphocyte (LYMP), monocyte (MON), granulocyte (GRAN), hemoglobin (HGB), and Hematocrit (Hct) Levels were evaluated.

Histopathological examination

The organs (brain, heart, thymus, lungs, liver, stomach, spleen, kidney, adrenal gland, pancreas and sex organs) excised from all the experimental rats were fixed in 10% formalin in labeled bottles, and processed for histological examination. Tissues embedded in paraffin wax were sectioned 5 mm thick and stained with haematoxylin and eosin, mounted on glass slides and examined under a standard light microscope⁽¹⁰⁾

Statistical analysis

All the data was expressed as Mean \pm SEM. Statistical analysis was tested by using One-way ANOVA followed by (Dunnett's test) using Graph pad prism version-8. The significance level was set at $P > 0.05$ for all tests. Group II, III, and IV will be statistically compared with Group I to find the treatment related effects.

RESULTS:

Chronic toxicity study

All the treatment group rats were administered with Urai mathirai drug solution at a dose of (10, 50 and 100 mg/kg b.wt) throughout the 90 days found to be no clinical toxicity signs such as physical observations, behavioral changes and other parameters such as body weight, food intake, water intake, respiration, convulsion, tremor, changes in eye and skin colors, etc were observed in the treated group compared to the control group. The observations were measured and summarized in Table 1.

Effect of Urai mathirai on relative organ body weight

The average and relative organ weight of Urai mathirai orally treated group of animals (at dose of 10, 50 and 100 mg/kg b.wt) and control groups showed statistically non-significant differences ($P > 0.05$). The results revealed that, the internal organs of rats were not adversely affected throughout the treatment. The effect of Urai mathirai on principal organ weights relative to body weight were presented in Table 2.

Effect of Urai mathirai on biochemical parameters

The results of the various biochemical parameters on the experimentally treated rats with the oral administration of the Urai mathirai at a dose of (10, 50 and 100 mg/kg b.wt) and normal groups showed statistically non-significant differences ($P > 0.05$). The results revealed that no abnormal changes in serum biochemical parameters such as albumin, total protein, globulin, Total bilirubin, urea, sodium, creatinine and uric acid levels etc., when compared to control group. The effect of Urai mathirai on biochemical parameters measured and summarized in Table 3.



Effect of Urai mathirai on Hematological parameters

The hematological parameters white blood cell (WBC), red blood cell (RBC), lymphocyte (LYMP), monocyte (MON), granulocyte (GRAN), hemoglobin (HGB), and Hematocrit (Hct) Levels were within normal limits compared to control group. No significant differences ($P > 0.05$) between treated animals with the Urai mathirai orally at dose of 10 mg/kg, 50 mg/kg and 100 mg/kg and control group rats were found. The hematological parameters were measured and summarized in Table 4.

Effect of Urai mathirai in Histopathological Study

Sections of lung, liver, kidney, spleen, heart, and brain tissues were perfused with 10% formalin and stored in the same and used for histopathological studies. The tissues were then embedded in molten paraffin wax. Sections were cut at 5 μ m thickness and stained with haematoxylin and eosin. The sections were then viewed under light microscope. The macroscopic examination of organs of treated rats revealed no abnormalities in the colour or texture when compared with the organs of the control group. Although some differences have been observed in the histopathological slides were presented in Figure 1-8.

Histopathology

Histopathological findings of lungs showed mild pulmonary oedema, multi focal mild mononuclear cell infiltration in peribronchiolar and alveolar region of control rats (Group I) and also treated rats groups (Group II to Group IV) of Urai mathirai orally at a dose level of (10 mg/kg, 50 mg/kg and 100 mg/kg). It revealed that no significant changes were observed related to drug toxicity effect and pictures were presented in the Figure 1.

Histopathological findings of liver showed sinusoidal congestion, very mild hepatocellular degeneration and focal hepatocellular necrosis in both control rats (Group I) and also in treated rats groups (Group II to Group IV) of Urai mathirai orally at a dose level of (10 mg/kg, 50 mg/kg and 100 mg/kg). It revealed that no significant changes were observed related to drug toxicity effect and pictures were presented in the Figure 2.

Histopathological findings of kidney showed mild tubular epithelial cell degeneration and focal mild interstitial mononuclear cell infiltration in both control rats (Group I) and also in treated rats groups (Group II to Group IV) of Urai mathirai orally at a dose level of (10 mg/kg, 50 mg/kg and 100 mg/kg). It revealed that no significant changes were observed related to drug toxicity effect and pictures were presented in the Figure 3.

Histopathological findings of spleen showed no abnormality in both control rats and also in treated rats groups (Group II to Group IV) of Urai mathirai orally at a dose level of (10 mg/kg, 50 mg/kg and 100 mg/kg). It revealed that no significant changes were observed related to drug toxicity effect and pictures were presented in the Figure 4.

Histopathological findings of heart showed no abnormality in both control rats and also in treated rats groups (Group II to Group IV) of Urai mathirai orally at a dose level of (10 mg/kg, 50 mg/kg and 100 mg/kg). It revealed that no significant changes were observed related to drug toxicity effect and pictures were presented in the Figure 5.

Histopathological findings of brain showed no abnormality in both control rats and also in treated rats groups (Group II to Group IV) of Urai mathirai orally at a dose level of (10 mg/kg, 50 mg/kg and 100 mg/kg). It revealed that no significant changes were observed related to drug toxicity effect and pictures were presented in the Figure 6.

Histopathological findings of glandular and non glandular stomach showed no abnormality in both control rats and also in treated rats groups (Group II to Group IV) of Urai mathirai orally at a dose level of (10 mg/kg, 50 mg/kg and 100 mg/kg). It revealed that no significant changes were observed related to drug toxicity effect and pictures were presented in the Figure 7.

A. Group I	B. Group II
C. Group III	D. Group IV

Figure 1 (A-D) Histopathological findings of Lungs in different groups of rats.

A. Group I	B. Group II
C. Group III	D. Group IV

Figure 2 (A-D) Histopathological findings of Liver in different groups of rats.

A. Group I	B. Group II
C. Group III	D. Group IV

Figure 3 (A-D) Histopathological findings of Kidney in different groups of rats.

A. Group I	B. Group II
C. Group III	D. Group IV

Figure 4 (A-D) Histopathological findings of Spleen in different groups of rats.

A. Group I	B. Group II
C. Group III	D. Group IV

Figure 5 (A-D) Histopathological findings of heart in different groups of rats.

A. Group I	B. Group II
C. Group III	D. Group IV

Figure 6 (A-D) Histopathological findings of brain in different groups of rats.

A. Group I	B. Group II
C. Group III	D. Group IV

Figure 7 (A-D) Histopathological findings of stomach in different groups of rats.



Histopathological findings of pancreas showed no abnormality in both control rats and also in treated rats groups (Group II to Group IV) of Urai mathirai orally at a dose level of (10 mg/kg, 50 mg/kg and 100 mg/kg). It revealed that no significant changes were observed related to drug toxicity effect and pictures were presented in the Figure 8.

A. Group I B. Group II
C. Group III D. Group IV

Figure 8 (A-D) Histopathological findings of pancreas in different groups of rats

CONCLUSION:

Urai mathirai administered in 3 doses by oral route (at therapeutic dose of 10 mg/kg, average dose 50 mg/kg and high dose 100mg/kg) regularly for 90 days did not produce any toxicity in animal models. There were no statistically significant alterations found in behavior, biochemistry, hematological parameters, organ weight and histopathology during the experimental period.

Table 1: General appearance and behavioral observations of control and treated groups.

Response	Group I (Normal)	Group II (10mg/kgb.w)	Group III (50mg/kgb.w)	Group IV (100mg/kgb.w)
Colour	Colour	Colour	Colour	Colour
Fur	N	N	N	N
Eyes	N	N	N	N
Mucous	N	N	N	N
Membrane				
Urine	N	N	N	N
Behavioral observations	Behavioral observations	Behavioral observations	Behavioral observations	Behavioral observations
Mood	N	N	N	N
CNS Excitation	NO	NO	NO	NO
CNS Depression	NO	NO	NO	NO
Motor Indication	Motor Indication	Motor Indication	Motor Indication	Motor Indication
Abnormal gait	NO	NO	NO	NO
Righting reflex	N	N	N	N
Posture	N	N	N	N
Sensory Responses	Sensory Responses	Sensory Responses	Sensory Responses	Sensory Responses
Touch & pain response	N	N	N	N
Straube's phenomenon	NO	NO	NO	NO
Reflexes	Reflexes	Reflexes	Reflexes	Reflexes
Pinna & corneal	N	N	N	N
Autonomic effects	Autonomic effects	Autonomic effects	Autonomic effects	Autonomic effects
Response	Group I (Normal)	Group II (10mg/kgb.w)	Group III (50mg/kgb.w)	Group IV (100mg/kgb.w)
Defecation & Lacrimation	N	N	N	N
Urination & Salivation	N	N	N	N
Piloerection	N	N	N	N
Miosis & Mydriasis	NO	NO	NO	NO
Diarrhoea	NO	NO	NO	NO
Respiratory effect	Respiratory effect	Respiratory effect	Respiratory effect	Respiratory effect
Apnoea & dyspnoea	NO	NO	NO	NO
Death	Death	Death	Death	Death
	NO	NO	NO	NO

N-Normal No-No Abnormality Observed



Table 2: Effect of Oral administration of Urai mathirai on Relative organs weight (g) of rats.

Organs	Group I (Normal)	Group II (10mg/kg.b.w)	Group III (50mg/kg.b.w)	Group IV (100mg/kg.b.w)
Brain	6.88±1.11	7.14±0.59	6.58±0.58	7.17±0.49
Heart	3.90±3.73	3.81±0.72	3.75±0.12	3.80±0.45
Thymus	3.41±1.83	2.23±1.11	0.75±0.14	0.89±0.19
Lungs	7.53±0.47	6.61±0.36	7.01±0.44	7.88±1.22
Liver	39.60±4.11	34.43±1.46	36.04±1.32	39.06±5.04
Stomach	6.14±0.86	6.48±0.42	5.73±0.18	6.60±0.45
Spleen	2.42±0.18	2.91±0.29	2.94±0.16	2.56±0.47
Pancreas	1.64±0.95	3.30±0.36	4.35±1.64	3.19±0.38
Kidney	8.26±1.05	8.55±0.96	7.92±0.19	8.53±0.78
Adrenal gland	0.34±1.20	0.27±0.07	0.20±0.04	0.22±0.05

All values are expressed as mean ± SEM (n=8).
No significant difference since $p > 0.05$, as compared to control group.

Table 3: Effect of oral administration Urai mathirai on biochemical parameters at the end of the treatment period

Organs	Group I (Normal)	Group II (10mg/kg.b.w)	Group III (50mg/kg.b.w)	Group IV (100mg/kg.b.w)
Blood glucose(mg/dl)	72.47±6.83	71.00±5.97	75.36±6.88	72.71±6.12
Total Cholesterol levels (mg/dl)	73.13±3.78	71.64±3.04	68.86±4.49	70.65±3.57
Triglyceride levels (mg/dl)	103.00±10.05	121.64±14.94	109.93±9.90	101.29±13.96
HDL levels (mg/dl)	30.87±6.70	25.07±1.41	25.36±1.76	23.47±1.26
LDL levels (mg/dl)	28.27±3.65	23.00±3.36	21.43±3.41	27.56±2.89
SGOT levels (U/L)	164.40±12.31	190.43±14.45	194.29±12.72	162.53±4.64
SGPT levels (U/L)	62.60±2.07	64.79±4.10	64.50±3.17	67.76±9.74
ALP levels (U/L)	200.60±19.15	183.21±16.86	220.36±20.74	165.47±14.62
GGT levels (U/L)	4.60±0.46	4.92±0.47	4.64±0.37	4.71±0.32
Total Protein levels (g/dl)	7.67±0.11	7.50±0.10	7.59±0.17	7.64±0.18
Albumin levels (g/dl)	3.41±0.07	3.44±0.10	3.37±0.13	3.45±0.06
LDH levels (U/L)	1234.13±65.36	1184.50±82.60	1232.50±91.74	14090.12±18.76
CRP levels (mg/L)	0.48±0.06	0.54±0.09	0.50±0.08	0.58±0.11
Creatinine Kinase levels (U/L)	814.87±70.65	904.79±124.68	887.07±139.57	651.65±41.45
Creatinine Kinase –MB levels (U/L)	507.80±43.57	496.50±50.51	446.14±36.92	369.76±18.04
Urea levels (mg/dl)	30.67±1.68	32.57±2.26	32.57±1.25	34.71±1.57
Serum creatinine levels (mg/dl)	0.53±0.02	0.53±0.02	0.56±0.02	0.54±0.03
Total Bilirubin levels (mg/dl)	0.12±0.01	0.11±0.01	0.11±0.01	0.10±0.00
Uric acid levels (mg/dl)	0.13±0.02	0.12±0.01	0.13±0.02	0.12±0.01
Serum Calcium levels (mg/dl)	9.98±0.12	9.89±0.12	9.12±0.68	14.22±4.49

All values are expressed as mean ± SEM (n=15, 14, 15, 17).
No significant difference since $p > 0.05$, as compared to control group.

Table 4: Effect of oral Urai mathirai on haematological parameters at the end of the treatment period

Parameters	Group I (Normal)	Group II (10 mg/kg b.w)	Group III (50 mg/kg b.w)	Group IV (10 mg/kg b.w)
WBC ($10^9/L$)	12.10±0.64	11.59±0.99	10.94 ±0.71	10.52±0.80
RBC ($10^{12}/L$)	7.42±0.17	7.78±0.31	7.66 ±0.21	7.76 ±0.24
LYMP ($10^9/L$)	8.13±0.40	9.36±1.10	7.54 ±0.55	7.11 ±0.55
MON ($10^9/L$)	0.31±0.02	0.31±0.03	0.32 ±0.03	0.31 ±0.03
GRAN ($10^9/L$)	3.68±0.28	3.21±0.38	3.08 ±0.24	2.81 ±0.29
HGB (g/dL)	12.00±0.27	12.44±0.47	12.19±0.26	19.61±7.41
HCT (%)	38.44±0.87	39.36±1.56	38.17±0.87	39.32±1.37

All values are expressed as mean ± SEM (n=13, 14, 15, 17).
No significant difference since $p > 0.05$, as compared to control group, except $p < 0.05$ in group,



In Vitro A-amylase inhibitory assay and Anti-oxidant, phytochemical screening of polyherbal siddha formulation Madhumega chooranam

INTRODUCTION:

Siddha System of Medicine is one of the age old medical system is being practiced in India. This system provides better solutions to manage many life style disorders. Type 2 Diabetes mellitus is one of the major disorder which impacts important threat due to its prevalence, chronicity and disabling complications. DM (Diabetes mellitus) is a metabolic disorder characterized by chronic hyperglycemia or increased a blood glucose level which affects the carbohydrate, fat and protein metabolism resulting from complete or relative lack of insulin secretion [1]. It prevalence increases globally and projected as 300 million by 2025. In India more than 10 million populations will suffer by this metabolic disorder [2]. Many therapeutic strategies are applied recently to observe the possible mechanism in type 2 diabetes mellitus [3]. The conventional method is α -amylase inhibition assay on cell lines. The objective of this study is to enumerate the basic pharmacological process of α -amylase inhibition assay on popularly known Siddha drug Madhumegachooranam[4] by cell line studies.



MATERIAL AND METHODS:

Siddha Medicine Madhumega chooranam(MMC) is purchased from a registered pharmacy. The chemicals used in the present study were of analytical grade and purchased from a reputed laboratory. The ethanolic extract was subjected into α -amylase inhibition by the method described by a standard method. The same extract has been tested for Anti oxidant activity involving different methods and phyto chemical analysis executed to assess its qualitative nature.



Preparation of MMC extract

The extracts are prepared by refluxing in ethanol for 72 hours followed by solvent recovery using rotary evaporator. The extracts are resuspended in 1% DMSO in a final concentration of 10mg/ml.

1. In vitro α -amylase inhibitory assay [5-6]

Principle:

The reducing sugars produced by the action of α -amylase react with dinitrosalicylic acid and reduce it to a brown coloured product, nitro amino salicylic acid.

Procedure:

Twenty five microlitres of 10 mg/ml extract and 25 μ l of 25mM phosphate buffer pH 6.9, containing porcine α -amylase at a concentration of 0.5 mg/ml were incubated at 25 $^{\circ}$ C for 10 min. After pre incubation, 25 μ l of 0.5% starch solution in 25mM phosphate buffer pH 6.9 was added. The reaction mixtures were then incubated at 25 $^{\circ}$ C for 10 min. The reaction was stopped with 50 μ l of 96mM 3, 5 dinitrosalicylic acid colour reagent. The micro plate was then incubated in a boiling water bath for 5 min and cooled to room temperature. Absorbance was measured at 540nm using a microplate reader.

Calculation:

$$\% \text{ inhibition} = \frac{\text{control test}}{\text{control}} \times 100$$

2. Determination of Anti oxidant activity

Anti oxidant activity of MMC extract is performed by using assay of Nitric oxide scavenging activity, Super oxide free radical scavenging activity and DPPH assay. The results were tabulated for further discussion.

a) DPPH Assay (2, 2-diphenyl -1-picrylhydrazyl) [7-8]

The radical scavenging activity of different extracts was determined by using DPPH assay according to a standard method. The decrease in the absorption of the DPPH solution after the addition of an antioxidant was measured at 517 nm. Ascorbic acid (10mg/ml DMSO) was used as reference.



Principle

1,1-diphenyl-2-picryl hydrazyl is a stable free radical with red colour which turns yellow when scavenged. The DPPH assay uses this character to show free radical scavenging activity. The scavenging reaction between (DPPH) and an antioxidant (H-A) can be written as,

$\text{DPPH} + [\text{H-A}] \rightarrow \text{DPPH-H} + (\text{A})$ Antioxidants react with DPPH and reduce it to DPPH-H and as consequence the absorbance decreases. The degree of discolouration indicates the scavenging potential of the antioxidant compounds or extracts in terms of hydrogen donating ability.

Reagent Preparation

0.1mM DPPH solution was prepared by dissolving 4mg of DPPH in 100ml of ethanol.

Procedure

Different volumes (1.25-10 μ l) of plant extracts were made up to 40 μ l with DMSO and 2.96ml DPPH (0.1mM) solution was added. The reaction mixture incubated in dark condition at room temperature for 20 minutes. After 20 minutes, the absorbance of the mixture was read at 517nm. 3ml of DPPH was taken as control.

Calculation:

$$\% \text{ inhibition} = \frac{\text{control test}}{\text{control}} \times 100$$

Nitric Oxide Scavenging Activity[9-11]

Nitric oxide (NO.) has also been involved in a variety of biological functions, including neurotransmission, vascular homeostasis, antimicrobial, and antitumor activities. Despite the possible beneficial effects of NO, its contribution to oxidative damage is also reported. This is due to the fact that NO can react with superoxide to form the peroxynitrite anion, which is a potential oxidant that can decompose to produce OH and NO. The procedure is based on the principle that, sodium nitro prusside in aqueous solution at physiological pH spontaneously generates nitric oxide which interacts with oxygen to produce nitrite ions that can be estimated using Griess reagent. Scavengers of nitric oxide compete with oxygen, leading to reduced production of nitrite ions. Large amounts of NO may lead to tissue damage. Nitric oxide scavenging activity was measured spectrophotometrically. Sodium nitro prusside (5mmolL⁻¹) in phosphate buffered saline pH 7.4, was mixed with different concentration of the extract (250-2500 μ g mL⁻¹) prepared in methanol and incubated at 25°C for 30minutes. A control without the test compound, but an equivalent amount of methanol was taken. After 30minutes, 1.5mL of the incubated solution was removed and diluted with 1.5mL of Griess reagent (1% sulphanilamide, 2% phosphoric acid and 0.1% N-1-naphthyl ethylene diaminedihydrochloride). Absorbance of the chromophore formed during diazotization of the nitrate with sulphanilamide and subsequent coupling with N-1 naphthyl ethylene diaminedihydrochloride was measured at 546nm and the percentage scavenging activity was measured with reference to the standard.

3. Phytochemical Analysis [12]

Phytochemicals, chemical compounds that occur naturally in plants (phyto means "plant" in Greek), are responsible for color and biological properties. The term is generally used to refer to those chemicals that may have biological significance but are not established as essential nutrients. The following tests are used for the analysis of phytochemicals as described by a standard method. The following tests are used for the analysis of phytochemicals present in the alcoholic extract of the tested drug. Some of the tests are done based on the standard procedure to assess the presence of alkaloid, flavanoid, phenols, glycosides, terpenoids and tannins. The inferences were listed in table 1.

Test for Alkaloids Dragandroff's test

8g of Bi(NO₃)₃. 5H₂O was dissolved in 20 ml HNO₃ and 2.72g of potassium iodide in 50 ml H₂O. These were mixed and allowed to stand. When KNO₃ crystals out, the supernatant was discarded off and made up to 100 ml with distilled water. The alkaloids were regenerated from the precipitate by treating with Na₂CO₃ followed by extraction of the liberated base with ether. To 0.5ml of alcoholic solution of extract added to 2.0 ml of HCl. To this acidic medium 1.0 ml of reagent was added. An orange red precipitate produced immediately indicates the presence of alkaloids.



Test for Flavanoids

Shinoda's test

In a test tube containing 0.5 ml of alcoholic extract 5-10 drops of dilute HCl and a small piece of ZnCl₂ or Mg were added and the solution was boiled for few minutes. In the presence of flavanoids reddish pink or dirty brown color was produced.

Test for Phenol

Ferric chloride test

To 2 ml of alcoholic solution of extract, 2 ml of distilled water followed by drops of 10% aqueous solution of FeCl₃ solution were added. Formation of blue or green indicates the presence of phenols.

Test for Glycosides

A small amount of alcoholic extract was dissolved in 1 ml of H₂O and the aqueous NaOH solution was added. Formation of yellow color indicates the presence of glycosides.

Test for Tannins

Ferric chloride test

To 1 -2 ml of aqueous extract, few drops of 5% aqueous ferric chloride solution was added. A bluish black colour, which disappears in addition of a few ml of sulfuric acid, formation of yellowish brown precipitate.

Test for Triterpenoids

LibermannBurchard test

The extract, 10 mg was dissolved in 1 ml of chloroform; 1 ml of acetic anhydride was added following the addition of 2 ml of Conc. H₂SO₄. Formation of reddish violet colour indicates the presence of triterpenoids.

Table 1. Phytochemical Analysis of Madhumega chooranam

Test	Observation	Inference
Alkaloids	Orange red precipitate was found	Presence of alkaloid(++)
Flavanoids	Pink colour was formed	Presence of Flavanoid(+)
Phenols	No characteristic change was observed	Absence of phenols
Glycosides	Yellow colour was formed	Presence of glycosides(++)
Terpenoids	Red colour was formed in the chloroform layer	Presence of terpenoids(++)
Tannins	No characteristic change was observed	Absence of tannins(-)

Statistical Analysis

Statistical difference and linear regression analysis were performed using Graphpad prism 6 statistical software.

RESULTS AND DISCUSSION:

The results showed that there was a concentration dependent α -amylase inhibitory effect of crude extract of Madhumegachooranam. At the concentration increased from 12.5 to 100 μ g/ml, percentage of inhibition increased from 37.22 % to 61.65 %. At a concentration of 100 μ g/ml there was a decrease in cell viability (Table 1, Figure 2&3). The IC₅₀ value was obtained at 20.2 μ g/ml (Figure 3). The antioxidant activity observed in different methodologies showed that Madhumegachooranam is having significant anti oxidant activity (Figure.3) Phytochemical analysis revealed that the presence of alkaloid, flavanoid, glycosides and terpenoids (Table 1). They are frequently implicated as having anti diabetic effect [13].

Antioxidants slow down the oxidative damage of our body. Antioxidants act as free radical scavengers. Preventing and repairing damages. Health problems such as Heart diseases, cancer and degenerative disorders are all exacerbated by oxidative damage. The Antioxidant activity of the drug was tested by DPPH and Nitric oxide scavenging activity.



Figure 2. In vitro α -amylase inhibition assay on Madhumega chooranam

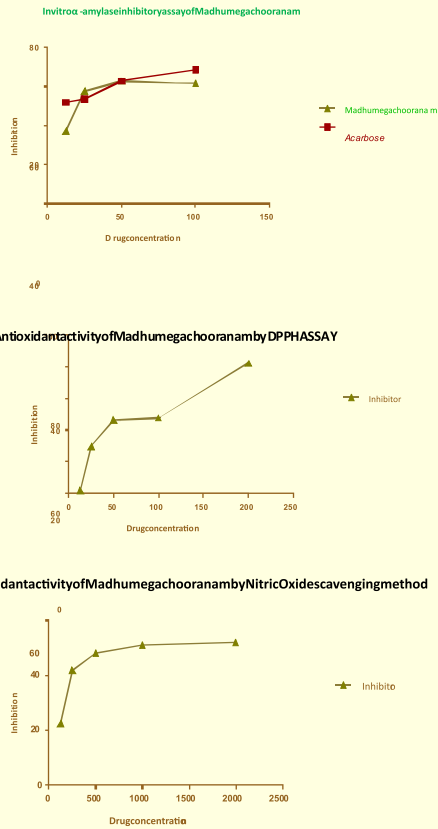


Figure 3. In vitro α -amylase inhibition and anti oxidant activity on Madhumegachooranam

CONCLUSION

It is concluded that the tested drug Madhumegachooranam exhibited significant α -amylase inhibition on dose dependant manner and having marked anti oxidant activity. The phyto constituents may help in restoring normal health. Thus this drug may act as a combined way in controlling hyperglycemia with minimal untoward effects. The drug could be tested for the maximum limit of controlling hyperglycemia in clinical studies. Further studies are needed to distinguish the pharmacological pathway in the insulin metabolism.

Ref: Ravichandran.M^{1*}, Mubarak.^{H2}

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TIP BITS: Guduchi (*Tinospora cordifolia*)

- Medhya** - The fresh juice of Gudūcī (20ml twice daily) may be used (C.S.Ci.1/1).
- Jvara** - (a) Gudūcī Svarasa (juice) + Satāvarī svarasa equal parts (10ml each) are mixed together and given along with jaggery (Guda), in vāta jvara (S.S.Ut. 39/174)
(b) Decoction prepared with Gudūcī, Parpati & Āmalakī (500- 100ml) may be administered in case of Pitta Jvara (Ha. Sam.3/ 2/71)
- Prameha** - Gudūcī Svarasa with honey (A.H.Ci.12/6)
- Ślīpada** - Gudūcī Svarasa along with gingelly oil (Taila) is given orally (C.D.42/16)
- Amlapitta** - Leaves of Gudūcī, Nimba and Patola are made into juice and administered along with honey (B. P.Ci.10/16)
- Chardi** - Gudūcī Hima Kaṣāya may be given orally along with honey (Bha.Pra.Ci. 17/21).
- Vatarakta** - Chronic administration of Guduchi in either juice or paste or powder or decoction form will definitely cure Vātarakta (B.P.Ci. 29/41 & V M 23/10)





10th WORLD AYURVEDA CONGRESS & AROGYA EXPO

DIGITAL HEALTH
AN AYURVEDA PERSPECTIVE

We had the privilege of participating as a proud sponsor in the 10th World Ayurveda Congress & Arogya Expo, held in Dehradun, Uttarakhand, from December 12 to 15, 2024. This prestigious event centered around the theme Digital Health: An Ayurveda Perspective.

As a leading manufacturer, SKM Siddha and Ayurveda showcased our latest innovations, including newly launched medicines such as Manyawin Forte, Katiwin, Gynedote, Ovorex, the Psorasiddh range, and more, at our vibrant Stall No. J2.

Our team of dedicated doctors and marketing professionals engaged with visitors, providing them with in-depth information about our products, ensuring they had direct access to the details they needed. It was a rewarding experience to connect with attendees and share our commitment to advancing health through Ayurveda.



National Siddha Day Celebration

On December 19, 2024, our company proudly celebrated National Siddha Day, commemorating the birth anniversary of Siddhar Agathiyar. The occasion was marked with traditional rituals, including a special pooja conducted to honor Siddhar Agathiyar's contributions to Siddha medicine. This celebration reflected our deep commitment to preserving and promoting the rich heritage of Siddha practices.



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